

Dansk Selskab for Flowcytometri

www.flowcytometri.dk

It is our pleasure to invite you to the

56th meeting of the Danish flow cytometry society (DSFCM)

"New Applications and Protocols in Flow Cytometry"

Date: 3. November 2016, kl 11:00-16:00 Location: Auditorium1, Aarhus Universitetshospital Tage-Hansens Gade 2, INDGANG 4A, 8000 Aarhus C,

Please see attached map

We have gathered vendors that will speak about new protocols and instruments in flow cytometry. The presentation will cover protocol and instrument related news from the companies that support the society.

Program: 11.00-11.15	Welcome
11.15-12.00	Session 1 DuraClone technology from Beckman Coulter
12.00-13.00	Lunch and exhibition
13.00-13.45	Session 2 Tips and tricks for extracellular vesicle analysis, Tina Van den Broeck from BD Biosciences
13.45-14.00	Break
14.00-14.45	Session 3 Spectral Technology in Flow Cytometry from Sony Biotechnology
14.45-15.15	Coffee and exhibition
15.15-16.00	Session 4 Flow Cytometer ZE5, Sebastian Hedlund from Bio-Rad Laboratories

Please visit <u>www.flowcytometri.dk</u> for updates on the program

Practical information:

- There will be served sandwiches in the lunch break and coffee and cake in the small afternoon break
- Cooperate members of the DSFCM have the possibility to present advertisement and products at a stand next to the auditorium. Please register via email to jpc@sund.ku.dk with "attending at 56th meeting" as headline no later than Tuesday 21.10.2016

Registration:

- All are welcome and the attendance is free of charge.
- However, registration is required since space is limited, and we need to order sandwich for you.
- Please register via email to jpc@sund.ku.dk with "56th dsfcm meeting" as headline no later than Tuesday 21.10.2016

Parking on campus is for payment, but the road next to Botanic Garden and The Old City has 5 hour parking.

Looking forward seeing you in Aarhus

On behalf of the board Jan P. Christensen

Abstracts:

Session 1:

DuraClone technology from Beckman Coulter, provide stable dry reagents that eliminates the need for manual reagent formulations or sustained cold chain storage, thereby substantially lowering variability coming in from operator or workflow.

Abstract: Multi parametric flow cytometry is a valuable tool for clinical research and requires use of multiple tandem dyes to monitor several markers simultaneously. Ensuring standardization in sample preparation, reagent handling and analysis methods is vital for clinical research studies (single or multicentric) that employ high throughput flow cytometry. The use of tandem dyes needs frequent re-compensation to account for changes in spectral spillover arising from the degradation of the tandem dye over time and manufacturing lot to lot variability in tandem dye preparations. These drawbacks along with liquid reagent handling reduce the overall robustness of the process as it may result in incorrectly compensated data, and manual variations in staining. DuraClone technology, providing stable dry reagents eliminates the need for manual reagent formulations or sustained cold chain storage, thereby substantially lowering variability coming in from operator or workflow.

Session 2:

Tips and tricks for extracellular vesicle analysis.

Submicron-sized extracellular vesicles (EVs) are released by virtually all cells and are more and more recognized as important players in intercellular communication, both in physiologic and pathophysiologic conditions. Flow cytometry is still one of the most commonly used techniques for EV analysis. However, since flow cytometers were primarily designed for cell analysis, the accurate measurement of EVs is challenged by their small size and low refractive index, which causes them to have scatter profiles similar to noise and plasma proteins. Due to these unique challenges, careful consideration should be given to instrument design, assay set-up and sample preparation prior to running a sample on a 'regular' flow cytometer.

Session 3:

Spectral Technology in Flow Cytometry

The Sony Spectral Analyzers use spectral technology to optimize sensitivity and enhances dim signal detection by collecting photons from 420nm to 800nm. Spectral technology also simplifies multicolor panel design, by eliminating band-pass filters and conventional compensation matrices to allow greater flexibility. This revolutionary approach uses spectral unmixing to expand the way cellular and microbiological samples are analyzed delivering the most accurate visualization of fluorescent populations available to scientists using flow cytometry. Spectral unmixing makes analysis simpler and easier by separating individual spectral fingerprints to allow scientists to better visualize their data. This delivers a more comprehensive picture to see rare populations and decreases the complexities associated with working with fluorescent proteins and multi laser excited fluorochromes. In this talk, we will discuss the Spectral Flow Cytometry in details and present cases that describe how experiments run on both Spectral Technology and conventional flow cytometry. These case studies will clarify conditions when data quality is advanced both in depth of information as well as eliminating bias data when analyzing cell populations and single cells using spectral flow cytometry.

Session 4:

Flow Cytometer ZE5 presentation from Bio-Rad Laboratories.

The ZE5 Cell Analyzer was designed from the ground up with user needs in mind. The integrated plate loader, with automated mixing and active temperature control, allows for switching from plate based analysis (96/384 shallow or deep) to tube based analysis (5ml/1.5ml) with zero effort. Innovative features like the EYE, provide automated feedback to users, letting them know if their filters are set up correctly, avoiding costly mistakes. On board QC allows for scheduled startup so the system is ready to go from the moment you arrive at the lab. The ZE5 also has bidirectional sample handling and absolute counting, without the limitations of a syringe based delivery system. Unused sample can be returned, and reagents, such as a viability dye, can be automatically added to your sample immediately before analysis. Configurable with up to 5 lasers and 28 colors, the ZE5 offers flexibility and power in an easy to use, compact format.

Aarhus Universitetshospital

Tage-Hansens Gade 2 - 8000 Aarhus C



Fælles AKUT Afdeling

AKUT afsnit 3 Indgang 1D

Senge- og behandlingsafd.		
Ortopædkirurgisk Afdeling	Ε	
ESA2 (E-elektiv sengeafsnit 2)	Indgang	1A
Ortopædkirurgisk Operationsafsnit E-OP	Indgang	2B
Ortopædkirurgisk Sekretariat	Indgang	1B
Knæ og Hofte Ambulatorium	Indgang	2B
Hånckirurgisk Ambulatorium	Indgang	1A
Skulder og Albue Ambulatorium	Indgang	1A
Skulder og Albue Sekretariat	Indgang	1A
Idrælsklinik	Indgang	2B
Knogleforskning	Indgang	9A
Kno cleforskning	Indgang	10/
Forskningskontorer	Indgang	7B
Forskningskontorer	Indgang	10A
Lægekontorer	Indgang	3C
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Geriatrisk Afdeling	G	
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Kirurgisk Afdeling Afsnit 260-280 Kirurgisk Sengeafsnit Kirurgisk Ambulatorium Indgang 1A Kirurgisk Dagafsnit Indgang 1A Kirurgisk Sengeafsnit 240 / Mammaendokrin Klinik (MEK) Indgang 1D Analfysiologisk- og Stomi-klinik Lægekontorer Forskningskontorer

Hæmatologisk Afdeling	R	
Afsnit 7-70-170 Hæmatologisk Sengeafsnit	Indgang	11A
Afsnit 7 Ambulatorium	Indgang	11A
Hæmatologisk Modtagelse	Indgang	1D
Hæmatologisk Ambulatorium	Indgang	4A
Hæmatologisk Ambulatorium R220 Amb	Indgang	1D
Hæmatologisk Undersøgelsesstue	Indgan g	1D
Immunhæmatologisk Laboratorium	Indgang	4A
Cancercytogenetisk Laboratorium	Indgan g	4A 11A
Hæmatologisk Sekretariat Lægekontorer	Indgang	14A
Hæmatologisk KFE	Indgan g Indgan g	3A
Personaleindgang	Indgang	1H
Psykolog	Indgang	3C
Projektsygeplejersker	Indgang	3C
Vagtværelse	Indgang	11A
"Hellen"	Indgang	15

Øvrige afdelinger m.v.

Anæstesiafsnit Sekretariat / Lægekontorer Apotekkontor Arkitekthuset	Indgang Indgang Indgang Indgang	2B 2G 3C 8A
AUDITORIER OG MØDELOKALER Auditorium 1	Indgang	4A
Auditorium 2-3-4-5 Undervisningslokale 1 Wødelokale 3 T-lokale 3	Indgang Indgang Indgang Indgang	13 <i>A</i> 3A 3A 2G
Centraldepot	Indgang	4A
Dagkirurgisk Afsnit	Indgang	2C
Driftafdelingen - syd	Indgang	3A
Dyrestald Ergo- og Fysioterapiafdeling	Indgang	9A
Sekretariat / Kontorer	Indgang Indgang	2B 2G
Forskningsbibliotek	Indgang	3C
HR Løn og personale	Indgang	3C
INFORMATION	Indgang	3B
IT-afdeling / Undervisning	Indgang	2G
Kantine	Indgang	4A
Kapel	Indgang	5A
Kapelassistent	Indgang	5E
Klinisk Biokemisk Afdeling	Indgang	2B
Ambulatorium	Indgang	2B 2A
BlodbankVareindevering	Indgang Indgang	2F
Køkken Vareindevering	Indgang Indgang	7C 7D
Linnedcentral	Indgang	4A
Medicoteknisk Laboratorium	Indgang	4A
Natindgang	Indgang	2E
Operationsgang / personaleindgang	Indgang	2G
OVITA (overvågning / intensivafsnit)	Indgang	2C
PATIENTHOTEL	Indgang	7A
Patologisk Afdeling	Indgang	5G
Portørvagten	Indgang	2B
Post	Indgang	4D
Præst	Indgang	10
Røntgen og Skanning. Lægekontorer	Indgang Indgang	2B 2F
Smerteteam	Indgang	2C
Snedkerværksted	Indgang	6D
Socialrådgiver	Indgang	10/
Teknisk Afdeling	Indgang	6A
Vareindevering	Indgang	6C 10 <i>A</i>
Uddannelsesansvarlige	Indgang	102
Undervisning / bioanalytikere	Indgang	5G
Vagtværelser	Indgang	4A
Vagtværelser	Indgang Indgang	7B
Vareindlevering (Centraldepot – vasketøj)	Indgang	4D
Varmtvandsbassin	Indgang	11/





